

East African Medical Journal Vol. 102. 10 October 2025

EVOLVING ANTIMICROBIAL RESISTANCE IN *VIBRIO CHOLERAE*: INSIGHTS FROM KENYA, 2015–2023

Waqo Gufu Boru, Department of Community Health, Epidemiology and Biostatistics, School of Public Health, Mount Kenya University, P.O. Box 342–01000, Thika, Kenya, Alfred Owino Odongo, Department of Community Health, Epidemiology and Biostatistics, School of Public Health, Mount Kenya University, P.O. Box 342–01000, Thika, Kenya, John Kariuki, Department of Community Health, Epidemiology and Biostatistics, School of Public Health, Mount Kenya University, P.O. Box 342–01000, Thika, Kenya

Corresponding author: Waqo Gufu Boru, Department of Community Health, Epidemiology and Biostatistics. School of Public Health, Mount Kenya University, P.O. Box 342–01000, Thika, Kenya. Email: waqogufu@gmail.com

## EVOLVING ANTIMICROBIAL RESISTANCE IN *VIBRIO CHOLERAE*: INSIGHTS FROM KENYA, 2015–2023

W. G. Boru, A. O. Odongo and J. Kariuki

### ABSTRACT

**Background:** Antimicrobial resistance (AMR) is a significant public health problem, with a considerable number of attributable and associated deaths both globally and in Kenya. There is an inherent data gap on AMR for *Vibrio cholerae* in Kenya. The present study aimed to document the pattern of Antimicrobial Resistance to *Vibrio cholerae* in Kenya.

**Methods:** This study used a retrospective, descriptive, and cross-sectional design. Data on the antimicrobial susceptibility and resistance of *V. cholerae* to various antibiotics were extracted from the National Public Health Laboratory (NPHL) of the Ministry of Health, Kenya, between 2015 and 2023. The data were subjected to descriptive analysis to determine the rates of sensitivity, intermediate resistance and antimicrobial resistance.

**Results:** The findings reveal that *V. cholerae* isolates were highly sensitive to chloramphenicol (94.0%), tetracycline (90.2%), gentamicin (86.8%), levofloxacin (94.1%), and cefuroxime (85.5%). Further, the isolates were highly resistant to Ampicillin (94.0%), Nalidixic acid (100%), Erythromycin (77.3%), and Ceftazidime (66.7%). Over two thirds (68.3%) of the isolates exhibited AMR, while 31.7% showed resistance to only one type of antibiotic. Among the isolates that exhibited resistance, 40.7% were resistant to two antibiotics, while 27.6% were resistant to more than two antibiotics. The predominant AMR pattern overall was Ciprofloxacin + Cefepime, while Ampicillin + Ciprofloxacin + Nalidixic acid was the predominant pattern of resistance to more than two antibiotics.

**Conclusion:** This study demonstrates an increasing trend of Antimicrobial Resistance (AMR) among *Vibrio cholerae* isolates in selected Kenyan counties, particularly to commonly used antibiotics, such as ampicillin, nalidixic acid,

and erythromycin. Conversely, high sensitivity was observed for chloramphenicol, levofloxacin, and tetracycline. Notably, a significant proportion of the isolates exhibited Multi Drug Resistance (MDR), posing a serious threat to effective cholera treatment. In order to mitigate this trend, routine surveillance of AMR in *V.cholerae* is recommended along with stringent antimicrobial stewardship.

**Keywords:** Antimicrobial Resistance, Sensitivity and *V. cholerae*

## INTRODUCTION

Antimicrobial resistance (AMR) is a significant public health problem worldwide. Over a Million deaths are associated with AMR, with 1.27 million deaths directly attributed to AMR. Evidence indicates that the misuse of antimicrobials in humans, animals, and plants is the main cause of AMR in bacterial strains <sup>1</sup>. In Africa, approximately 1.05 million deaths in 2019 were associated with AMR, with lower respiratory and thoracic infections accounting for the highest percentage, followed by bloodstream and abdominal infections <sup>2</sup>. According to a review by the UK government, AMR is projected to kill 10 million people annually by 2050 <sup>3,4</sup>.

Evidence indicates that bacteria cause AMR through biochemical mechanisms, such as destruction or enzymatic modification of antibiotics, modification of antibiotic target sites, decrease in antibiotic penetration, and elimination of antibiotics from the cell by efflux pumps <sup>5</sup>. Based on this evidence, governments and agencies in the health sector and researchers agree that AMR is a significant problem that requires urgent, globally coordinated action plans to deal with <sup>6,7</sup>. Resistance to commonly prescribed antimicrobials is a major threat to the fight

against cholera and other endemic bacterial diseases in Africa and worldwide<sup>8</sup>. Microorganisms that have been documented to exhibit resistance include *Escherichia coli*, *Klebsiella pneumonia*, *Neisseria spp*, *Salmonella serovars*, and *Vibrio cholerae* <sup>9</sup>.

*Vibrio cholerae* O1 and O139 serogroups that exhibit AMR are associated with the majority of global cholera outbreaks<sup>10,11</sup>. In low-middle-income countries such as Kenya, there has been growing concern about the emergence of AMR *V. cholerae* strains<sup>12</sup>. Evidence suggests that AMR *V. cholerae* is resistant to the majority of antibiotics used in treatment, which is exhibited by considerably high treatment failures of the disease<sup>13</sup>. Multi Drug Resistance (MDR) enteric pathogens were first discovered in the 1960s <sup>14,15</sup>. In the past few decades, AMR in *V. cholerae* has mutated rapidly and spread globally, owing to the overuse and misuse of antibiotics in different sectors. Extensive drug resistance (EDR) and MDR *V. cholerae* genes are highly enriched with mobile genetic elements (MGEs) and can horizontally transfer resistance genes to other pathogenic microorganisms <sup>12</sup>.

According to World Health Organization (WHO), there are limited publications and reports on AMR in Africa, including Kenya

(9). Evidence further suggests the need for the development of comprehensive and expounded data on AMR in *V. cholerae*<sup>16</sup>. In Kenya, AMR remains a significant health problem. For instance, in 2019, 8500 deaths were directly attributable to AMR, and another 37,300 deaths were associated with AMR<sup>17</sup>. During the recent cholera epidemics in Kenya, new *V. cholerae* strains exhibiting new resistance have emerged. Cholera is endemic to Kenya, and in the past few decades, the country has experienced three major protracted epidemics. However, there are considerable limitations to the data and information on the AMR patterns of *V. cholerae* at the national level<sup>18</sup>. This study aimed to document the AMR of *V. cholera* isolates in Kenya.

## Methodology

### Study design

This study adopted a retrospective descriptive design, assessing the susceptibility and resistance of *V. cholerae* clinical isolates.

### Study area

A total of 279 isolates from 14 counties in Kenya between 2015 and 2023 were considered in the present study. The counties were selected based on the burden of cholera outbreak. The 14 counties exhibited high burden of cholera outbreak during the study period. The counties were Nairobi, Lamu, Machakos, Mandera, the Tana River, Kajiado, Murang'a, Kiambu, Kitui, Garissa, Turkana, Isiolo, and Narok.

### Study period

The considered study period was between 2015 and 2023.

### Cholera data source

Data on susceptibility and resistance to *V. cholerae* on various antibiotics were extracted from the National Public Health Laboratory (NPHL) of the Ministry of Health, Kenya. The AMR data for *V. cholerae* was consolidated between 2015 and 2023.

### Data analysis

Descriptive analysis was performed using SPSS version 25, and bar graphs were drawn using RStudio 4.3.3. The data are presented in the tables and graphs as provided in the results section.

### Ethical consideration

To ensure data privacy and protection, no personal identifiers including names, addresses and contact information or any other directly identifying information were collected or extracted from the secondary data sources. Ethical approval and authorization to conduct this study were obtained from the Mount Kenya University Ethical Review Committee (Ref: MKU/ISERC/4306) and the National Commission for Science, Technology, and Innovation (NACOSTI) (License no: NACOSTI/P/24/39710). Approval to access Cholera AMR data was obtained from the Kenya Ministry of Health Division of National Public Health Laboratory (NPHL).

## Results

### Socio-demographic characteristics

This study included 279 cases from 14 counties in Kenya, covering the period

between 2015 and 2023 (Table 1). The majority (44.8%) of the samples were from Nairobi County, followed by Lamu County, and the least were from Isiolo and Narok Counties. The majority (65.9%) of the

samples were from male patients, whereas the least (30.5%) were from female patients. The mean age of the cholera patients was  $33.73 \pm 15.94$  years.

**Table 1: Socio-demographic characteristics of the cholera cases (n=279)**

| Variable           | Frequency | Percentage (%) |
|--------------------|-----------|----------------|
| <b>Gender</b>      |           |                |
| Male               | 184       | 65.9           |
| Female             | 95        | 34.1           |
| <b>Age (years)</b> |           |                |
| $33.73 \pm 15.94$  |           |                |
| <b>County</b>      |           |                |
| Nairobi            | 158       | 56.6           |
| Lamu               | 48        | 17.2           |
| Machakos           | 18        | 6.5            |
| Manderra           | 13        | 4.7            |
| Tana River         | 13        | 4.7            |
| Kajiado            | 10        | 3.6            |
| Murang'a           | 7         | 2.5            |
| Kiambu             | 3         | 1.1            |
| Kitui              | 3         | 1.1            |
| Garissa            | 2         | 0.7            |
| Turkana            | 2         | 0.7            |
| Isiolo             | 1         | 0.4            |
| Narok              | 1         | 0.4            |

#### **Antimicrobial susceptibility of *V. cholerae***

More than three-quarters of the isolates were sensitive to chloramphenicol (94.0%), gentamicin (86.8%), cefuroxime (85.5%), tetracycline (90.2%), and levofloxacin (94.1%) (Table 2). Majority of the isolates were sensitive to ciprofloxacin (57.9%), amikacin (53.5%), and cefotaxime (56.0%). A large number of the isolates were resistant to

ampicillin (94.0%), all the isolates were resistant to nalidixic acid (100%), erythromycin (77.3%), and ceftazidime (66.7%). Additionally, over half (61.8%, 57.7%, 57.7%, and 51.1%) of the isolates were resistant to trimethoprim-sulfamethoxazole, meropenem, ceftazidime, and cefepime, respectively. However, only few (6) isolates were subjected to Nalidixic acid.

**Table 2: Antimicrobial susceptibility of *Vibrio cholerae* (n=279)**

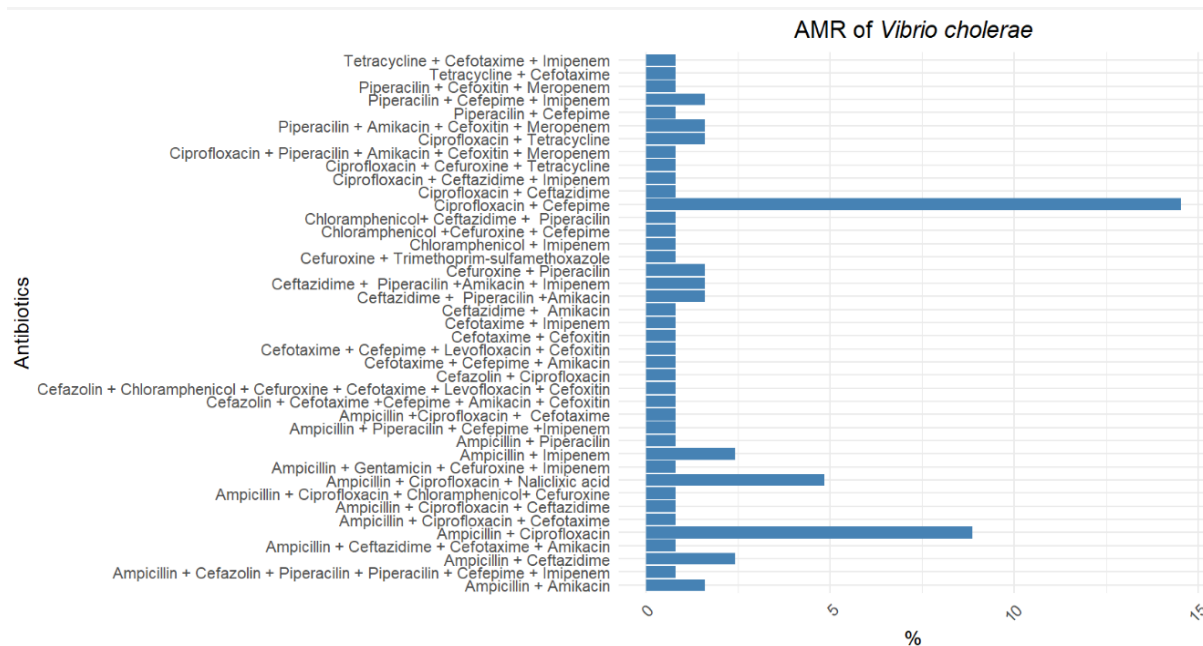
| Antimicrobial Susceptibility Testing |                   |                      |                    |
|--------------------------------------|-------------------|----------------------|--------------------|
| Antibiotic type                      | Sensitive<br>f(%) | Intermediate<br>f(%) | Resistance<br>f(%) |
| Ampicillin                           | 3(6.0)            | (0)                  | 47(94.0)           |
| Ciprofloxacin                        | 73(57.9)          | 1(0.8)               | 52(41.3)           |
| Chloramphenicol                      | 79(94.0)          | (0)                  | 5(6.0)             |
| Gentamicin                           | 33(86.8)          | 3(7.9)               | 2(5.3)             |
| Cefuroxime                           | 53(85.5)          | (0)                  | 9(14.5)            |
| Tetracycline                         | 83(90.2)          | (0)                  | 9(9.8)             |
| Cefepime                             | 17(37.8)          | 5(11.1)              | 23(51.1)           |
| Piperacillin                         | 21(47.7)          | 2(4.5)               | 21(47.7)           |
| Amikacin                             | 23(53.5)          | 6(14.0)              | 14(32.6)           |
| Cefotaxime                           | 28(56.0)          | 7(14.0)              | 15(30.0)           |
| Levofloxacin                         | 32(94.1)          | 0(0)                 | 2(5.9)             |
| Ceftazidime                          | 14(33.3)          |                      | 28(66.7)           |
| Erythromycin                         | 5(22.7)           |                      | 17(77.3)           |
| Trimethoprim-sulfamethoxazole        | 13(38.2)          |                      | 21(61.8)           |
| Nalidixic acid                       | 0(0)              | 0(0)                 | 6(100)             |
| Meropenem                            | 2(28.6)           | 1(14.3)              | 4(57.1)            |
| Cefoxitin                            | 3(21.4)           | 3(21.4)              | 8(57.1)            |

Key: f=frequency, % = Percentage

### Anti-Microbial Resistance (AMR) of *Vibrio cholerae*

As shown in figure 1 more than two-thirds of the isolates (68.3%) exhibited resistance while 31.7% exhibited resistance to only one antibiotic. The most predominant AMR pattern was Ciprofloxacin + Cefepime (14.6%) followed by Ampicillin + Ciprofloxacin (8.9%) and then Ampicillin + Ciprofloxacin + Nalidixic acid (4.9%). Of the 68.3% isolates that exhibited AMR, 40.7% exhibiting resistance to two antibiotics and 27.6% exhibiting resistance to more than two

antibiotics. The predominant pattern of resistance to two antibiotics in the isolates was Ciprofloxacin + Cefepime, followed by Ampicillin + Ciprofloxacin and then Ampicillin + Ceftazidime. For the isolates that exhibited resistance to more than two antibiotics the predominant pattern was Ampicillin + Ciprofloxacin + Nalidixic acid, followed by piperacillin + amikacin + cefoxitin + meropenem, ceftazidime + piperacillin + amikacin + imipenem, piperacillin + cefepime + imipenem, and ceftazidime + piperacillin + amikacin.



**Figure 1: AMR of *Vibrio cholerae***

## Discussion

The study findings indicated that cholera was more common among males. This finding was similar to a study done in Kenya, that investigated cholera epidemics from 2007 to 2010 and 2015 to 2016, which documented that the incidence of cholera was higher in males than in females<sup>18</sup>. This observation could potentially be attributed to men having poor health seeking behavior, high tendency of eating and drinking out and working in unhygienic environments which increases risk of exposure<sup>19,20</sup>. Most isolates demonstrated a high sensitivity to Chloramphenicol, Gentamicin, Cefuroxime, Tetracycline, and Levofloxacin, indicating that these antibiotics are effective treatment options. Isolates from the 2007 to 2010 cholera epidemic in Kenya, cholera outbreaks in Kisumu County in 2014, and cholera outbreaks in Uganda in 2017 showed high sensitivity to Chloramphenicol, Tetracycline, and Gentamicin, respectively<sup>18,21,22</sup>. Furthermore, a Cuban study reported high sensitivity of *V. cholerae* isolates to chloramphenicol (92.0%) and tetracycline (91.2%)<sup>23</sup>. Studies from East Delhi and

Mozambique have documented the sensitivity of *V. cholerae* to chloramphenicol (24,25). A meta-analysis of the global antimicrobial status of *V. cholerae* identified Gentamicin and Cefotaxime to be highly effective against *V. cholerae*<sup>16</sup>. Additionally, more than half of the isolates were sensitive to Ciprofloxacin, Amikacin, and Cefotaxime, suggesting emerging resistance to these antibiotics, which may limit their clinical efficacy. Similarly, other studies conducted in Haiti, the United States, and France have reported low resistance of *V. cholerae* to Ciprofloxacin and Cefotaxime<sup>26,28</sup>.

The antimicrobial resistance of *V. cholerae* isolates has been increasing in recent years and has become a major risk to cholera treatment strategies<sup>29</sup>. The study revealed high rates of resistance to ampicillin (94.0%), nalidixic acid (100%), erythromycin (77.3%), and ceftazidime (66.7%). The high resistance to first-line antibiotics indicates that these antibiotics are ineffective. Similarly, Kenyan studies reported high resistance of *V. cholerae* to ampicillin (64%), trimethoprim-sulfamethoxazole (54%), and nalidixic acid (100%)<sup>30</sup>. The Ugandan and Cuban Studies also reported 100% and 60% resistance,

respectively, of *V. cholerae* isolates to ampicillin<sup>22,23</sup>. Additionally, isolates from the 2012 to 2015 Mozambique cholera outbreak exhibited 100% resistance to Ampicillin and Nalidixic acid<sup>31</sup>. More than half of the isolates were resistant to trimethoprim-sulfamethoxazole, meropenem, cefoxitin, and cefepime. Similarly, studies in Zambia, Democratic Republic of Congo (DRC) and North India have documented considerable medium resistance of *V. cholerae* to trimethoprim-sulfamethoxazole (SXT)<sup>32,34</sup>.

The study findings indicate the levels of resistance, particularly to important classes of antibiotics such as fluoroquinolones (ciprofloxacin and nalidixic acid), extended-spectrum cephalosporins (Cefepime and Ceftazidime), carbapenems (meropenem and imipenem), and penicillins (ampicillin and piperacillin). The high rates of resistance to multiple drug classes, especially carbapenems, which are often considered last-resort antibiotics, are particularly worrying from a clinical perspective. Furthermore, *V. cholerae* resistance to erythromycin is worrying considering that it is safe antibiotic of choice in pregnancy.

In the present study, two thirds (68.3%) of the isolates demonstrated MDR, with the most predominant pattern being Ciprofloxacin + Cefepime. The study findings align with a previous Kenyan study which reported an MDR rate of 64.5% in *V. cholerae* isolates from 2007 to 2010 and 2015–2016 cholera epidemics<sup>18</sup>. Similarly, studies from Ghana and South Africa also highlighted high MDR rates reporting over 97% and 36-50% MDR rates in clinical isolates<sup>38,39</sup>. In contrast a Bangladeshi study conducted among children and older adults documented a lower MDR rate of 28.1%, with the most predominant pattern being TMP-SMX+erythromycin+ampicillin<sup>35</sup>.

MDR *V. cholerae* isolates are not a new phenomenon, isolates resistant to

Ampicillin, Nalidixic acid, chloramphenicol, and tetracycline began appearing in the 1990s<sup>36</sup>. More recently, a 2021 Kenyan study detected and characterized MDR *V. cholerae* by using genes encoding Extended-Spectrum Beta-Lactamase (ESBL) enzymes. ESBL isolates are resistant to chloramphenicol, streptomycin, sulfamethoxazole, trimethoprim, tetracycline, rifampicin, and erythromycin<sup>8</sup>.

Evidence increasingly suggests an increase in MDR *V. cholerae* in Kenya and the region<sup>22,37</sup>. The proliferation of MDR among *V. cholerae* isolates as reported by a study in South African may be attributed to spontaneous mutations or horizontal transfer of resistance genes through the co-existence of gut coliforms and *Vibrio species*<sup>40</sup>. This widespread increase in antibiotic resistance majorly influenced by the extensive use of antibiotics has led to the emergence of MDR strains, hence posing a significant global public health concern<sup>18</sup>. Indeed, recent trends indicate that most clinical *V. cholerae* isolates are resistant to almost all routinely used antibiotics<sup>12</sup>.

### Limitations of the study

Key limitation of this study is the incomplete or inconsistent surveillance data from some counties in Kenya, especially in earlier years, which may compromise the accuracy and reliability of the resistance trend analysis. Additionally, limited geographic coverage, focusing only on selected countries, may restrict the generalizability of the findings to a broader national context. This study also faces the challenge of retrospective design constraints, as it relies on secondary data collected between 2015 and 2023, limiting control over potential confounding variables

and possibly missing emerging resistance patterns.

### Conclusion

In conclusion, the antimicrobial resistance profiles of the *V. cholerae* isolates from Kenya have significant clinical implications. Although there were high susceptibility rates to chloramphenicol, gentamicin, cefuroxime, tetracycline, and levofloxacin, alarming resistance to critical and common antibiotics was observed, including complete resistance to nalidixic acid and high resistance to ampicillin, erythromycin, and ceftazidime. The *V. cholerae* isolates also exhibited considerably high dual resistance and AMR. The emergence of resistance to carbapenems and the predominant MDR pattern involving ampicillin, ciprofloxacin, and nalidixic acid, which significantly limits treatment options for cholera, indicate the need for enhanced surveillance and development of strategies to address the issue of MDR in cholera management in Kenya.

### Recommendation

There is need for evaluation of the observed high resistance of the commonly prescribed antibiotics namely; Nalidixic acid and erythromycin among the Cholera patients. Need for strengthening surveillance of Antimicrobial Resistance in *V. Cholerae* during Cholera outbreak

### Acknowledgment

The authors express their gratitude to the research participants for their enthusiastic participation.

Author contributions: Conceptualization; W.G, AO and JK. Methodology; All authors. Validation; All authors; Data collection; W.G. Data curation; W.G. Formal analysis and drafting paper; W.G. Review and Editing of the manuscript; All authors.

### Conflict of interest statement

The author declares that there is no conflict of interest.

### Source of funding

The research was self-funded.

### Data availability statement

The data supporting the findings of this study are available from the corresponding author upon reasonable request. The AMR data for cholera were obtained from National Public Health Laboratory (NPHLS) records, with approval from the Ministry of Health, Kenya. Due to the sensitive nature of the data and ethical considerations, data cannot be shared publicly but may be made available to qualified researchers upon request and with appropriate institutional and ethical approvals.

### References

1. World health organization. *Antimicrobial resistance* [internet]. 2023 [cited 2025 Aug 11]. Available from: <https://www.who.int/news-room/fact-sheets/detail/antimicrobial-resistance>
2. Sartorius B, Gray AP, Davis Weaver N, Robles Aguilar G, Swetschinski LR, Ikuta KS, et al. The burden of bacterial antimicrobial resistance in the WHO African region in 2019: a cross-country systematic analysis. *Lancet Glob Heal*. 2024;12(2):e201–16.
3. O'Neill J. Antimicrobial Resistance: Tackling a crisis for the health and wealth of nations. *Afr J Lab Med*. 2014;7(2):1-12.
4. O'Neill J. *Tackling drug-resistant infections globally: final report and recommendations* [Internet]. London (UK): Review on Antimicrobial Resistance; 2016 [cited 2025 Sep 07]. Available from: [https://amr-review.org/sites/default/files/160518\\_Final%20paper\\_with%20cover.pdf](https://amr-review.org/sites/default/files/160518_Final%20paper_with%20cover.pdf)

5. Sefton AM. Mechanisms of antimicrobial resistance: Their clinical relevance in the new millennium. *Drugs*. 2002;62(4):557–66.
6. Centers for Disease Control and prevention (CDC). *antibiotic resistance threats in the United States, 2019* [Internet]. Atlanta (GA): U.S. department of health and human services, CDC; 2019 [cited 2024 Jul 06]. Available from: <https://www.cdc.gov/antimicrobial-resistance/data-research/threats/index.html>.
7. Wagenlehner FME, Dittmar F. Re: Global Burden of Bacterial Antimicrobial Resistance in 2019: A Systematic Analysis. *Eur Urol*. 2022;82(6):658.
8. Kariuki S, Wairimu C, Mbae C. Antimicrobial Resistance in Endemic Enteric Infections in Kenya and the Region, and Efforts Toward Addressing the Challenges. *J Infect Dis*. 2021;224(Suppl 7):S883–9.
9. World Health Organization, Regional Office for Africa. *Antimicrobial resistance in the WHO African Region: a systematic literature review* [Internet]. Brazzaville (CG): WHO Regional Office for Africa; 2021 [cited 2024 Jun 11]. Available from: <https://www.afro.who.int/publications/antimicrobial-resistance-who-african-region-systematic-literature-review>
10. Verma J, Bag S, Saha B, Kumar P, Ghosh TS, Dayal M, et al. Genomic plasticity associated with antimicrobial resistance in *Vibrio cholerae*. *Proc Natl Acad Sci U S A*. 2019;116(13):6226–31.
11. Ramamurthy T, Mutreja A, Weill FX, Das B, Ghosh A, Nair GB. Revisiting the Global Epidemiology of Cholera in Conjunction With the Genomics of *Vibrio cholerae*. *Front Public Heal*. 2019;7(July):1–10.
12. Das B, Verma J, Kumar P, Ghosh A, Ramamurthy T. Antibiotic resistance in *Vibrio cholerae*: understanding the ecology of resistance genes and mechanisms. *Vaccine*. 2020;38:83–92.
13. Clemens JD, Nair GB, Ahmed T, Qadri F, Holmgren J. Cholera. *Lancet*. 2017;390(10101):1539–49.
14. Watanabe T. Infective Heredity of Multiple Drug Resistance in Bacteria. *Bacteriol Rev*. 1963;27(1):87–115.
15. Nakaya R, Nakamura A, Murata Y. Resistance transfer agents in *Shigella*. *Biochem Biophys Res Commun*. 1960;3(6):654–9.
16. Yuan X hui, Li Y mei, Vaziri AZ, Kaviar VH, Jin Y, Jin Y, et al. Global status of antimicrobial resistance among environmental isolates of *Vibrio cholerae* O1/O139: a systematic review and meta-analysis. *Antimicrob Resist Infect Control* [Internet]. 2022;11(1):1–11. Available from: <https://doi.org/10.1186/s13756-022-01100-3>
17. Institute for Health Metrics and Evaluation (IHME). *The burden of antimicrobial resistance in Lebanon* [Internet]. Seattle (WA): IHME, University of Washington; 2019 [cited 2025 Jun 21]. Available from: [https://www.healthdata.org/sites/default/files/files/Projects/GRAM/Lebanon\\_0.pdf](https://www.healthdata.org/sites/default/files/files/Projects/GRAM/Lebanon_0.pdf)
18. Shah MM, Bundi M, Kathiiko C, Guyo S, Galata A, Miringu G, et al. Antibiotic-Resistant *Vibrio cholerae* O1 and Its SXT Elements Associated with Two Cholera Epidemics in Kenya in 2007 to 2010 and 2015 to 2016. *Am Soc Microbiol*. 2023;11(3):1–10.
19. Ngwa MC, Young A, Liang S, Blackburn J, Mouhaman A, Morris JG. Cultural influences behind cholera transmission in the far north region, republic of Cameroon: A field experience and implications for operational level planning of interventions. *Pan Afr Med J*. 2017;28:1–10.
20. Mageto LM, Mutono N, Aboje G, Gathura P, Okunga E, Muange A, et al. Spatio-temporal pattern and risk factors associated with cholera outbreaks in selected high-risk areas of Kenya. *Trans R Soc Trop*

- Med Hyg [Internet]. 2025;119(March):758–66. Available from: <https://doi.org/10.1093/trstmh/traf032>
21. Awuor SO, Omwenga EO, Daud II. Geographical distribution and antibiotics susceptibility patterns of toxigenic *Vibrio cholerae* isolates from. African J Prim Heal Care Fam Med. 2020;12(1):1–6.
22. Iramiot JS, Rwego IB, Kansiime C, Asiimwe BB. Epidemiology and antibiotic susceptibility of *Vibrio cholerae* associated with the 2017 outbreak in Kasese district, Uganda. BMC Public Health. 2019;19(1):1–9.
23. Fernández-Abreu A, Bravo-Fariñas L, Rivero-Navea G, Cabrera-Cantelar N, Nuñez-Fernández FA, Cruz-Infante Y, et al. Determinants of virulence and antimicrobial susceptibility in non-O1, Non-O139 *Vibrio cholerae* Isolates. MEDICC Rev. 2017;19(4):21–5.
24. Gujral L, Sema C, Rebaudet S, Taibo CLA, Manjate AA, Piarroux R, et al. Cholera epidemiology in mozambique using national surveillance data. J Infect Dis. 2013;208(SUPPL. 1):107–14.
25. Das S, Choudhry S, Saha R, Ramachandran VG, Sarkar BL. Brief Original Article Emergence of multiple drug resistance *Vibrio cholerae* O1 in East Delhi. J Infect Dev Ctries. 2011;5(4):294–8.
26. Baron S, Lesne J, Jouy E, Larvor E, Kempf I, Boncy J, et al. Antimicrobial susceptibility of autochthonous aquatic *Vibrio cholerae* in Haiti. Front Microbiol. 2016;7(OCT):1–12.
27. Baron S, Larvor E, Chevalier S, Jouy E, Kempf I, Granier SA, et al. Antimicrobial susceptibility among urban wastewater and wild shellfish isolates of Non-O1/Non-O139 *Vibrio cholerae* from La Rance estuary (Brittany, France). Front Microbiol. 2017;8(SEP):1–15.
28. Ceccarelli D, Chen A, Hasan NA, Rashed SM, Huq A, Colwell RR. Non-O1/Non-O139 *Vibrio cholerae* carrying multiple virulence factors and *V. cholerae* O1 in the Chesapeake Bay, Maryland. Appl Environ Microbiol. 2015;81(6):1909–18.
29. Das B, Verma J, Kumar P, Ghosh A, Ramamurthy T. Antibiotic resistance in *Vibrio cholerae*: Understanding the ecology of resistance genes and mechanisms. Vaccine [Internet]. 2020;38:A83–92. Available from: <https://doi.org/10.1016/j.vaccine.2019.06.031>
30. Kiiru S, Maina J, Mwaniki JN, Songoro E, Kariuki S. Enteric bacterial agents associated with diarrhea and their antimicrobial sensitivity profiles in children under 5 years from mukuru informal settlement, Nairobi, Kenya. BMC Infect Dis. 2024;24(17):1–10.
31. Dengo-Baloi LC, Semá-Baltazar CA, Manhique LV, Chitio JE, Inguane DL, Langa JP. Antibiotics resistance in El Tor *Vibrio cholerae* O1 isolated during cholera outbreaks in Mozambique from 2012 to 2015. PLoS One. 2017;12(8):1–10.
32. Mwansa JCL, Mwaba J, Lukwesa C, Bhuiyan NA, Ansaruzzaman M, Ramamurthy T, et al. Multiply antibiotic-resistant *Vibrio cholerae* O1 biotype El Tor strains emerge during cholera outbreaks in Zambia. Epidemiol Infect. 2007;135(5):847–53.
33. Berthe M, Sandra M, Muyembe JJ, George NT, Ickel Kakongo K, Daniel Yassa N. Antimicrobial Drug Resistance of *Vibrio cholerae*, Democratic Republic of the Congo. Emerg Infect Dis [Internet]. 2015;21(5):2015. Available from: <https://www.britannica.com/place/Democratic-Republic-of-the-Congo>
34. Mala E, Oberoi A, Alexander VS. *Vibrio* isolates from cases of acute diarrhea and their antimicrobial susceptibility pattern in a tertiary care hospital. Int J Basic Appl Sci. 2014;3(1):35–7.
35. Garbern SC, Chu TC, Yang P, Gainey M, Nasrin S, Kanekar S, et al. Clinical and socio-environmental determinants of multidrug-resistant *Vibrio cholerae* O1 in older children

- and adults in Bangladesh. *Int J Infect Dis*. 2021;105:436–41.
36. Yu L, Zhou Y, Wang R, Lou J, Zhang L, Li J, et al. Multiple antibiotic resistance of *Vibrio cholerae* serogroup O139 in China from 1993 to 2009. *PLoS One*. 2012;7(6):1–9.
37. Bundi M, Shah MM, Odoyo E, Kathiiko C, Wandera E, Miring'u G, et al. Characterization of *Vibrio cholerae* O1 isolates responsible for cholera outbreaks in Kenya between 1975 and 2017. *Microbiol Immunol*. 2019;63(9):350–8.
38. Igere BE, Okoh AI, Nwodo UU. Antibiotic susceptibility testing (AST) reports: A basis for environmental/epidemiological surveillance and infection control amongst environmental *Vibrio cholerae*. *Int J Environ Res Public Health*. 2020;17(16):1–23.
39. Abana D, Gyamfi E, Dogbe M, Opoku G, Opare D, Boateng G, et al. Investigating the virulence genes and antibiotic susceptibility patterns of *Vibrio cholerae* O1 in environmental and clinical isolates in Accra, Ghana. *BMC Infect Dis*. 2019;19(1):1–10.
40. Wang R, Li J, Kan B. Sequences of a co-existing SXT element, a chromosomal integron (CI) and an IncA/C plasmid and their roles in multidrug resistance in a *Vibrio cholerae* O1 El Tor strain. *Int J Antimicrob Agents* [Internet]. 2016;48(3):305–9. Available from: <http://dx.doi.org/10.1016/j.ijantimicag.2016.05.020>